

## D-6-Phosphogluconate

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### Principle

6-Phosphogluconic dehydrogenase (6-PG-DH) catalyses the oxidation of 6-phosphogluconate (gluconic acid-6-phosphate) by triphosphopyridine nucleotide (TPN):



The equilibrium of the reaction lies far to the right, so that with a slight excess of TPN and at pH 7–8, a quantitative oxidation of 6-phosphogluconate is obtained<sup>1)</sup>. The reaction proceeds very quickly. The increase of optical density at 334, 340 or 366 m $\mu$  due to the reduction of TPN is a measure of the reaction.

As it is not possible at present \*) to obtain a 6-phosphogluconic dehydrogenase preparation of sufficient purity (in particular, free from hexokinase and glucose-6-phosphate dehydrogenase), the enzymatic determination of 6-phosphogluconate can only be carried out on solutions which do not contain either glucose or glucose-6-phosphate. The method therefore cannot be used for tissue extracts.

### Reagents

1. Triethanolamine hydrochloride
2. Sodium hydroxide, A. R., 2 N
3. Magnesium chloride, A. R., MgCl<sub>2</sub>·6 H<sub>2</sub>O
4. Triphosphopyridine nucleotide, TPN  
monosodium salt, TPN·NaH<sub>2</sub>; commercial preparation, see p. 1029.
5. 6-Phosphogluconic dehydrogenase, 6-PG-DH  
from yeast, commercial preparation, see p. 993.

### Purity of the enzyme preparation

6-Phosphogluconic dehydrogenase is prepared from brewer's yeast according to<sup>1)</sup>. From 120 g. dried yeast about  $6.2 \times 10^3$  units \*\*) 6-PG-DH is obtained with a specific activity of 54 units/mg. The preparation can be freeze-dried without any great loss of activity (yield ca. 80%) and in this state it is stable for months. The preparation is contaminated with about 20% glucose-6-phosphate dehydrogenase and about 5% hexokinase (relative to the 6-PG-DH activity).

### Preparation of Solutions

Prepare all solutions with fresh, doubly distilled water.

- I. Triethanolamine buffer (0.4 M; pH 7.6):  
Dissolve 18.6 g. triethanolamine hydrochloride in about 200 ml. doubly distilled water, add 18 ml. 2 N NaOH and dilute to 250 ml.
- II. Magnesium chloride (0.1 M):  
Dissolve 2 g. MgCl<sub>2</sub>·6 H<sub>2</sub>O in doubly distilled water and make up to 100 ml.
- III. Triphosphopyridine nucleotide (ca.  $2 \times 10^{-2}$  M  $\beta$ -TPN):  
Dissolve 20 mg. TPN·NaH<sub>2</sub> in 1 ml. doubly distilled water.

\*) The enzyme has now been crystallized: *S. Pontremdi, A. de Flora, E. Grazi, G. Mangiarotti, A. Bonsignore and B. L. Horecker*, J. biol. Chemistry 236, 2975 [1961].

\*\*) According to *Bücher et al.* Definition, see p. 33.

<sup>1)</sup> *B. L. Horecker and P. Z. Smyrniotis*, J. biol. Chemistry 193, 371 [1951].

#### IV. 6-Phosphogluconic dehydrogenase, 6-PG-DH (ca. 800 units<sup>\*)</sup>/ ml.):

Take up the lyophilized preparation (about 16 mg. protein) in 1 ml. M/25 glycylglycine buffer pH 7.5 and remove the insoluble material by centrifuging in the cold.

##### Stability of the solutions

Store all the solutions, stoppered, in a refrigerator at 0–4°C. The enzyme solution is stable only for a few days. Preferably prepare the daily requirement of the enzyme by dissolving the lyophilized preparation freshly each time.

##### Procedure

##### Spectrophotometric measurements

*Preliminary remarks:* The concentration of 6-phosphogluconate in the assay mixture should not exceed  $1 \times 10^{-7}$  moles/ml. A blank can usually be omitted. The sample must be free from glucose-6-phosphate and fructose-6-phosphate, and its pH should be between 5 and 9.

*Method:* Wavelength: 340, 334 or 366 m $\mu$ ; light path: 1 cm.; final volume: 1.08 ml. Read against the control cuvette. Pipette successively into the cuvettes:

<i>Experimental cuvette</i>	<i>Control cuvette</i>
up to 0.7 ml. sample	2 ml. buffer (solution I)
make up to 1 ml. with buffer (solution I)	
0.01 ml. TPN solution (III)	
0.05 ml. MgCl <sub>2</sub> solution (II).	

Mix thoroughly, bring the contents of the cuvettes to room temperature and read the optical density  $E_1$  twice with an interval of 3 min. Mix into the experimental cuvette

0.02 ml. 6-PG-DH solution (IV).

When the optical density no longer increases (5–10 min. after the addition of the enzyme, depending on the 6-phosphogluconate concentration) read the optical density  $E_2$  twice with an interval of 3 min. The changes within 3 min. in the initial optical density  $E_1$  and the final optical density  $E_2$  are usually small in comparison to difference in optical density  $\Delta E = E_2 - E_1$  and can be neglected. If necessary, apply a correction (see p. 39). The correct functioning of the assay can easily be checked, if on completion of the reaction 0.010 ml.  $2 \times 10^{-3}$  M 6-phosphogluconate solution is added to the experimental cuvette. The increase in optical density should be complete in 3–5 min. Read the optical density  $E_3$  twice within 3 min. and calculate the difference  $\Delta E' = E_3 - E_2$ .  $\Delta E'$  should be 0.115 at 340 m $\mu$ , 0.112 at 334 m $\mu$  and 0.061 at 366 m $\mu$ .

##### Calculations

Under the stated conditions 6-phosphogluconate is quantitatively oxidized, so that the 6-phosphogluconate content of the sample can be calculated from the optical density difference  $\Delta E$ :

$$\frac{\Delta E \times V}{\epsilon \times d \times V_s} = \mu\text{moles 6-phosphogluconate/ml. sample.}$$

where  $V$  = assay volume in ml.  
 $V_s$  = volume of the sample in ml.  
 $d$  = light path of the cuvette in cm.  
 $E$  = optical density difference [ $E_2 - E_1$ ]  
 $\epsilon$  = extinction coefficient [cm.<sup>2</sup>/ $\mu$ mole].

<sup>\*)</sup> See footnote on p. 143.

If the measurements are made at 366 m $\mu$  then the contents of the cuvette must be equilibrated at 25°C, because the extinction coefficient is dependent on temperature at this wavelength<sup>2)</sup>.

**Example**

A 6-phosphogluconate solution (0.4 ml.) was tested and the measurements were made at 340 m $\mu$ :

Before addition of 6-PG-DH    0 min.  $E_1 = 0.068$

  3 min.  $E'_1 = 0.068$

After addition of 6-PG-DH    0 min.  $E_2 = 0.493$

  3 min.  $E'_2 = 0.493$

$$\Delta E = E_2 - E_1 = 0.425$$

$$\frac{0.425 \times 1.08}{6.28 \times 1 \times 0.4} = 0.183 \text{ } \mu\text{moles 6-phosphogluconate/ml. sample}$$

**Specificity**

In the absence of glucose, glucose-6-phosphate and fructose-6-phosphate the assay is specific for 6-phosphogluconate. Gluconate does not react.

<sup>2)</sup> *H. J. Hohorst, Biochem. Z. 328, 509 [1957].*